



Chronic Obstructive Lung Disease, Stem Cells and Telocytes: Review of Therapeutic

Idiberto José Zotarelli Filho^{1-3*}, Aldemir Bilaqui¹, Elias Naim Kassis², Marinônio Lopes Cornélio³ and Luiz Fernando Frascino¹

¹Portuguese Beneficent Hospital, São Jose do Rio Preto SP, Brazil

²Unipos - Post graduate and continuing education, Street Ipiranga, 3460, São José do Rio Preto SP, Brazil 15020-040

³State University of Sao Paulo-IBILCE-UNESP, Rua Cristovao Colombo 2265, Sao Jose do Rio Preto SP, Brazil

*Corresponding author: Dr. Idiberto Jose Zotarelli Filho, University of Sao Paulo-IBILCE-UNESP, Rua Cristovao Colombo 2265, Sao Jose do Rio Preto SP, Brazil, Tel: 11- 5627-0233; E-mail: m.zotarelli@gmail.com

Rec date: Jul 21, 2014 Acc Date: Mar 13, 2015 Pub Date: Mar 20, 2015

Abstract

Chronic Obstructive Lung Disease (COLD) is responsible for leading causes of mortality and morbidity worldwide, and a major public health problem worldwide. In addition, COLD is associated with high costs. In the EU, the total direct costs of respiratory diseases are estimated at 6 % of total health care costs, and that COLD accounts for 56 % of these expenditures (38.6 billion euros). In the United States, the direct costs of COLD are 29.5 billion and indirect costs of the order of 20.4 billion dollars. In Brazil, it is estimated that 5 million Brazilians have COLD, according to the Brazilian Thoracic Society (BTS) and second DATASUS, COLD generated in 2010, the National Public Health System, 141,994 hospitalizations that led the 778,428 days of hospitalization. Cell therapy can be regarded in a broad and general manner to the use of cells for the treatment of COLD, mainly stem cells and telocytes. Although not a significant number of reports, involving lung and object of study in cell therapy in human patients, there is consistent data, both in humans and in animal models that show the migration of stem cells from bone marrow to the lung and the recovery of lung function in different experimental situations. The results, together with those of other research groups, allow to contemplate the possibility of application of cell therapy in patients with COLD.

Keywords: COLD; Stemcells; Telocytes; Cell therapy

Introduction

Epidemiologic data of Chronic Obstructive Lung Disease

Chronic Obstructive Lung Disease (COLD) is responsible for leading causes of mortality and morbidity worldwide, and a major public health problem worldwide. In 2000, according to the World Health Organization - WHO, COPD was the 5th leading cause of mortality worldwide, with 4.5% of deaths, second only in brain and heart, AIDS or AIDS-vascular diseases (Acquired Immunodeficiency Syndrome) [1-3]. In 2004, also according to the WHO, COPD became the 4th leading cause of death worldwide, with 5.1% of deaths, surpassing AIDS [4-7]. Moreover, when we consider the "burden of

disease" (Burden of Disease) measured in QALYs (Quality Adjusted Life Years - metric involving the quantity and quality of years lived), it is estimated that COLD will be the 13th pathology Ranking world in 2004 to 5th in 2030, possibly due to population aging and the industrialization of low-income countries [7-9].

In addition, COLD is associated with high costs. In the EU, the total direct costs of respiratory diseases are estimated at 6% of total health care costs, and that COLD accounts for 56% of these expenditures (38.6 billion euros). In the United States, the direct costs of COLD are 29.5 billion and indirect costs of the order of 20.4 billion dollars [10]. In Brazil, it is estimated that 5 million Brazilians have COLD, according to the Brazilian Thoracic Society (BTS) and second DATASUS, COLD generated in 2010, the National Public Health System, 141,994 hospitalizations that led the 778,428 days of hospitalization [11]. The total cost of these hospitalizations was R \$ 92,434,415.51 and 7,937 deaths directly related to COLD. According to BTA, the prevalence of COLD in Brazil is 15.8% in adults older than 40 years [12].

The main issue related to COLD is cigarette smoking, which are associated with 95% of cases. Other causative factors include air pollution, occupational exposure to pollutants and also in alpha-1-antitrypsin [13-16] disability. In the epidemiological study called Proyecto Latinoamericano de Investigacion en pulmonary obstruction (PLATINUM, Latin American Project for the Investigation of Pulmonary Obstruction), designed for screening for COLD and conducted in São Paulo, it was found that 15.8% of individuals aged 40 years or older had COLD and that 12.5% of these individuals had never been exposed to tobacco smoke [17-21]. From these data, it appears that other risk factors for COLD than smoking, are important in Brazil, including AAT deficiency. A study in COLD patients present severe AAT deficiency in 2-3% of patients. According to the PLATINO study, there are 5 to 7 million with COLD in Brazil [22-26]. It is estimated. Epidemiological studies conducted worldwide have shown that AAT deficiency is about as frequent as cystic fibrosis, affecting one in every 2,000-5,000 individuals [27,28].

Obstructive diseases have high prevalence in humans and have been the subject of many clinical and experimental studies aimed at a broader understanding of its pathogenesis, pathophysiology, and especially the establishment of more rational forms of treatment [22-26]. As a result of this great effort, the concepts of obstructive diseases suffered an extraordinary expansion in recent years, involving the integration of mechanical factors, inflammatory agents, autonomic regulation of airways and environmental aspects [21-26].

According to GOLD 2011 the main aspects of therapy of patients with COLD are: the abandonment of smoking in smokers, which may be obtained either by frequent counseling patients, as by therapeutic measures [28-31]. Smoking cessation is the measure with the greatest potential to prevent disease progression [22-24].

Significance

Clinical standard treatment

According to the "Global Initiative for Chronic Obstructive Lung Disease" (GOLD), COLD is a common, preventable and treatable disease, which is characterized by persistent limitation in lung air flow, usually progressive and associated with character with increased

inflammatory response of the airways and lungs to noxious particles and gases [31-38]. Exacerbations and comorbidities contribute to the overall disease severity [39-42].

Moreover, the chronic airflow limitation characteristic of GOLD is caused by a mixture of small airway disease (obstructive bronchiolitis) and parenchymal destruction (emphysema), and the contribution of each of these processes varies from individual to individual [43-51]. Chronic inflammation causes structural changes and narrowing of small airways [51-54]. The destruction of the lung parenchyma, also by inflammatory processes, leads to a loss of adhesion of the small airways and alveoli pathways decrease the elasticity of these pathways, which consequently leads to a decreased ability of these remain open during expiration [55-59]. The airflow limitation is best measured by spirometry, and this is the most available and reproducible tool to assess lung function test [60-65].

Many previous definitions of GOLD have emphasized the terms "emphysema" and "chronic bronchitis", which are not included in the definitions used in GOLD. Emphysema, or destruction of the surface of pulmonary gas exchange (alveoli), is a term that is often pathological (but incorrectly) and used clinically only describes one of several structural abnormalities present in patients with GOLD [66-70]. Chronic bronchitis, or the presence of productive cough for at least 3 months a year for two consecutive years, remains a term clinically and epidemiologically useful [71-73]. However, it is important to recognize that chronic productive cough (chronic bronchitis) is an independent disease entity that may precede or follow the development of airflow limitation and may be associated with the development or acceleration of chronic airflow limitations [74-78].

Currently, GOLD is classified into 4 stages of severity depending on the degree of airway obstruction as measured by forced expiratory volume in 1 second (FEV1) and the ratio of FEV1 and forced vital capacity (FVC) - FEV1/FVC. These stages are listed below: Mild FEV1/FVC < 0.70; FEV1 ≥ 80% predicted, moderate FEV1/FVC < 0.70; 50% ≤ FEV1 < 80% predicted, severe FEV1/FVC < 0.70; 30% ≤ FEV1 < 50% predicted, very serious FEV1/FVC < 0.70; FEV1 < 30% predicted or FEV1 < 50% predicted plus chronic respiratory failure [79-81].

However, more recently these stages began to be called "degrees" and are used together with the level of symptoms presented by patients to classify them into four categories (A, B, C and D). According to the GOLD 2011, the purpose of the evaluation is to determine the GOLD disease severity, its impact on patient health and the risk of future events (exacerbations, hospital admission and death) in order to guide therapy. Thus, the following aspects of the disease are used separately: symptoms, degree of airflow limitation (using spirometry), exacerbation risk, comorbidities [82-91].

Typical symptoms of GOLD usually associated with hypersecretion of mucus and cough with fibrosis due to the attempted repair of chronic inflammation, obstruction of small airways because the lumen is reduced due to the formation of inflammatory exudate or by deposition collagen and emphysema with destruction of parenchyma, enlargement of the air, space narrowing and weakening of small airways with increased lymphoid follicles, loss of lung elasticity and closure of small airways. These presentations can be concurrent or not, and may show a variable degree [92-97].

Further, as treatment of GOLD, vaccination against influenza and pneumococcal infections that decrease the executives responsible for periods of exacerbation. Measures of rehabilitation and physical conditioning that improve the quality of life of patients and their

ability to perform physical activities. Using both drugs in the stable phase of the disease in order to decrease the intensity and frequency of seizures, the seizures in aiming the removal of the patient in this phase of acute exacerbation of the disease [76,77]. The pharmacotherapy used in the treatment of GOLD consists of several classes of medications as: Beta 2 agonists, short and long action (LABA), anticholinergics short and long action (LAMA), systemic and inhaled corticosteroids (ICS), the nonspecific phosphodiesterase inhibitors (methylxanthines) specific phosphodiesterase 4 inhibitors (roflumilast), as well as various combinations of such drugs, according to the degree of the disease [77].

Cell Therapy – Advancements Stem Cells and Telocytes

Often cases of GOLD are refractory to pharmacological treatments, then in order to meet such emergencies mortalities and morbidities and although cells, tissues and organs damaged by trauma or disease being recovered spontaneously by the body, the cell therapy regenerative medicine seeking to replace cells or tissues with severe injuries or senescent cells to regenerate and restore their functions [77,78]. The diseases that are targets of these treatments are the main causes of morbidity and mortality worldwide companies, taking as examples heart disease, diabetes, cancer, lung diseases, and genetic diseases.

Transplantation of hematopoietic adult stem cells in the bone marrow is still today the most used treatment for treatment of human stem cells and consists of the infusion of these cells to restore hematopoiesis in the recipient or even provide a greater quantity stem cells for promote tissue regeneration. The transplanted stem cells survive for long time maintaining their properties, being used for neoplastic and non-neoplastic diseases such as aplastic anemia, autoimmune diseases and lung diseases [70-79].

There is evidence that these cells can also transdifferentiate into other non-hematopoietic lineages such as endothelial and lung cells [70-82]. In addition to the hematopoietic stem cells in the bone marrow, there are mesenchymal stem cells around 0.01% to 0.001% which are multipotent cells capable of differentiating into various cells and tissues, increasing the therapeutic capacity of hematopoietic stem cells [80-82]. Despite a relatively small number of reported cases and the lung as an object of study in cell therapy, there is consistent data, both in humans and in animal models, which show the presence of stem cells labeled in the bone marrow lungs of animals or human patients undergoing bone marrow transplantation (Figure 1).

Others authors obtained promising results with regard to cell therapy in GOLD [35]. These authors transplanted adherent cells derived from bone marrow of transgenic mice in wild-type mice subjected to lung injury by bleomycin and verified the presence of transplanted cells as type I pneumocytes in the lung of the recipient. Krause et al. 2001 [36], after irradiation of a female, transplanted stem cells from the bone marrow of a male mouse and observed the presence of these stem cells with Y chromosomes in several tissues, including the lung tissue.

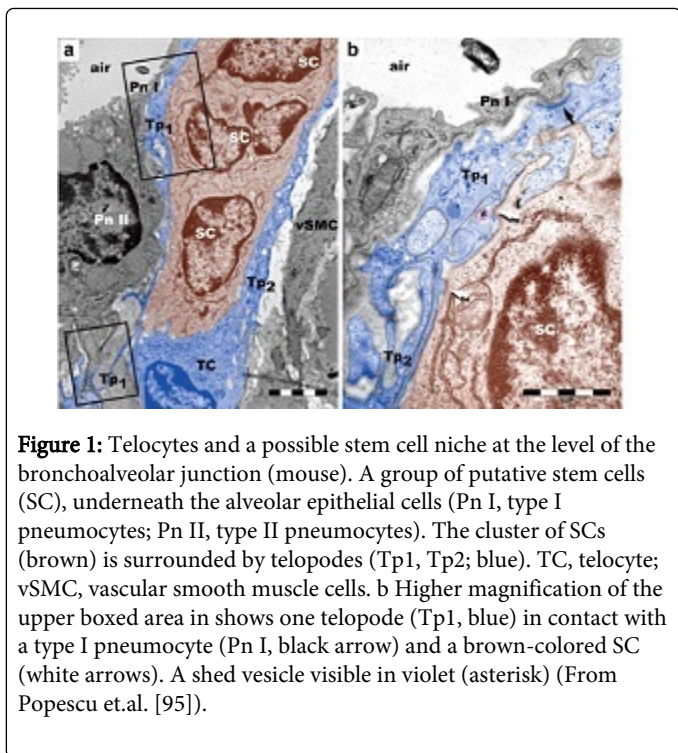


Figure 1: Telocytes and a possible stem cell niche at the level of the bronchoalveolar junction (mouse). A group of putative stem cells (SC), underneath the alveolar epithelial cells (Pn I, type I pneumocytes; Pn II, type II pneumocytes). The cluster of SCs (brown) is surrounded by telopodes (Tp1, Tp2; blue). TC, telocyte; vSMC, vascular smooth muscle cells. b Higher magnification of the upper boxed area in shows one telopode (Tp1, blue) in contact with a type I pneumocyte (Pn I, black arrow) and a brown-colored SC (white arrows). A shed vesicle visible in violet (asterisk) (From Popescu et.al. [95]).

non-cavitary organs (lung, trachea, pleural, skeletal muscle, exocrine pancreas, placenta, mammary gland and the [104-110]).

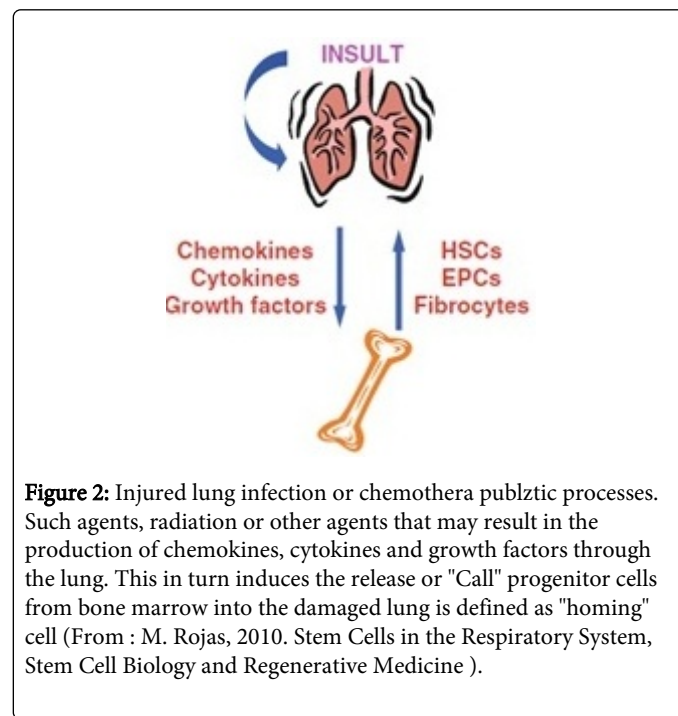


Figure 2: Injured lung infection or chemotherapy processes. Such agents, radiation or other agents that may result in the production of chemokines, cytokines and growth factors through the lung. This in turn induces the release or "Call" progenitor cells from bone marrow into the damaged lung is defined as "homing" cell (From : M. Rojas, 2010. Stem Cells in the Respiratory System, Stem Cell Biology and Regenerative Medicine).

Suratt et al. [71] selected lungs of women who have previously received allogeneic hematopoietic stem cell donor men and performed in situ hybridization studies, verifying the presence of the Y chromosome and endothelial and epithelial markers in these cells from the donors. Therefore found chimerism in human lung equivalent to 2.5 to 8% epithelial and 37.5 to 42.3% endothelial. Next, Ishizawa et al. 2004 [33] demonstrated, in elastase-induced emphysema model, treatment with retinoic acid and colony-stimulating factor granulocyte (G-CSF) leads to the regeneration with the presence of alveolar bone marrow cells in the lung after treatment, indicating the mobilizing these cells could be of great importance in regeneration.

Also using the lung as an organ of interest, Yamada et al. [81] lung injury induced by lipopolysaccharide derived from *Escherichia coli* and analyzed the mobilization of bone marrow progenitor cells, finding a large number of receptors in donor cells with damaged lungs. However, there is no standardized therapeutic effective of these cells, with a stratification results inconclusive answers. You need to know and establish the appropriate niche for biological treatment of COLD and control variables of the clinical conditions of each patient to ensure the therapeutic effective of these cells (Figure 2).

Added to this, the author Popescu et al. [98-103] first demonstrated the importance of the presence of Telocytes(TC) with stem cells to increase the efficiency of cell therapy. The Telocytes is a new type of interstitial cells of mesenchymal origin, first described in 2005 by Popescu et al. Initially, they were named as Cajal interstitial cells, but when the lack of resemblance to the canonical Cajal interstitial cells became obvious, the name Telocytes was introduced and is currently used.

The presence of TC has been documented in several organs and cavities in connective tissues (heart, the pericardium, the pulmonary vein, intestine, mesentery, gallbladder, uterus and fallopian tubes and

These are characterized by being very large, typically tens of micrometers, around a hundred micrometers or more. They are made by alternating dilated portions, called podoms and podomers. Podomers the thickness is usually very thin, below the resolving power of conventional light microscopy in sections, with the branch dichotomous extensive testing and immunohistochemistry with positive reactions for c-kit, vimentin and CD34 [111-113].

Apart from the conventional role of mechanical support for the TC network, we believe that the main role of TC is intercellular communication and regulation, with the existence of TP 3D network [114,115]. Thus, it is clear the location of TC and TP in close proximity to blood capillaries or nerve endings. We conclude that in adulthood, most of the TC has the fibroblastoid [104]. Immunocytochemical assays suggest that a variety of TC populations show different immunophenotypes may exist. Several papers were hypothesized to TC as mechanical function, intercellular signaling, guide immature cells during organogenesis, are precursors of many mesenchymal cells in adulthood [105-107]. However, none of these papers were also tested. Even so, it is proposed that TC can be acutely important in regeneration of tissues and organs [105,116-118].

These results, however, have been viewed with greater caution by some authors to question the accuracy of detection techniques employed [105,116-118]. The TC migration to the lung could have been overestimated, and therefore would be present in a much lower proportion than has been proposed, with questionable clinical significance [119,120]. Results in favor of this proposition were obtained by Xu et al. [83].

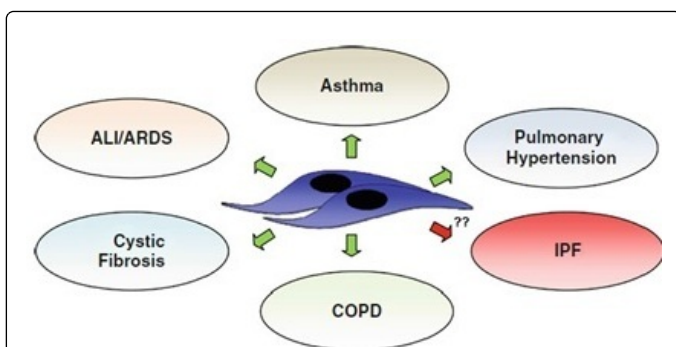


Figure 3: Action of mesenchymal stem cells (MSCs) in inflammatory lung diseases. MSCs are emerging as a therapeutic modality in many inflammatory lung diseases such as acute lung injury, chronic obstructive pulmonary disease, cystic fibrosis, pulmonary hypertension, and allergic diseases such as asthma. Due to its potential antifibrotic effects in vivo, MSC cannot represent a therapeutic option in patients with fibrotic lung diseases such as idiopathic pulmonary fibrosis (From: M. Rojas, 2010. Stem Cells in the Respiratory System, Stem Cell Biology and Regenerative Medicine).

These authors, after transplantation of mesenchymal stem cells in mice that had previously received intraperitoneal injection of lipopolysaccharide (LPS), not found in the immune histochemical analysis of lung tissue taken 14 days after transplantation, significant occurrence of cells with positive signal for mesenchymal stem cell - MSC [100, 104]. However, although they have not found evidence of effective integration of MSC to the lung tissue and the presence of derived cells with the phenotype of local cell types, the authors were able to prove that transplantation of stem cells has allowed a reduction in inflammation and pulmonary edema induced by LPS [120](Figure 3).

Conclusion

It was concluded that makes it important to adopt cell therapy as an alternative to conventional treatments for COLD, with the application of mesenchymal stem cells or hematopoietic stem cell associated with telocitos to better recovery of patients, emphasizing the improvement of quality of life.

Disclosure of potential conflicts of interest

The authors declare that they have no conflicts of interests.

Acknowledgements

We appreciate the support of Duke University (Durham, NC, USA) in the field of research and statistical studies and also appreciate the support of the Life Group (Brazil), Biosafe (Switzerland, EC) and Celartia (Ohio, USA), Portuguese Beneficent Hospital (Sao Jose of Rio Preto - SP, Brazil and Unipos - Postgraduate and continuing education LTDA (Sao Jose of Rio Preto- SP, Brazil).

References

- Abbott OA, Hopkins WA, Van Fleit WE, Robinson JS (1953) A new approach to pulmonary emphysema. *Thorax* 8: 116-132.
- Barnes PJ (2000) Chronic obstructive pulmonary disease. *N Engl J Med* 343: 269-280.
- Barnes PJ, Shapiro SD, Pauwels RA (2003) Chronic obstructive pulmonary disease: molecular and cellular mechanisms. *Eur Respir J* 22: 672-688.
- Barnes PJ (2003) New concepts in chronic obstructive pulmonary disease. *Annu Rev Med* 54: 113-129.
- Bast A, Haenen GR, Doelman CJ (1991) Oxidants and antioxidants: state of the art. *Am J Med* 91: 2S-13S.
- Bianco P, Robey P G (2000) Marrow stromal stem cells. *The Journal of Clinical Investigation*. 105: 1.163-1.668.
- Bittmann I, Dose T, Baretton GB, Müller C, Schwaiblmair M, et al. (2001) Cellular chimerism of the lung after transplantation. An interphase cytogenetic study. *Am J Clin Pathol* 115: 525-533.
- Böyum A (1968) Separation of leukocytes from blood and bone marrow. Introduction. *Scand J Clin Lab Invest Suppl* 97: 7.
- Brantigan O C, Mueller E, Kress M B (1959) A surgical approach to pulmonary emphysema. *Am Rev Respir Dis* 80: 194-206.
- Campos H S (2003) O preço da DPOC. *Pulmão R J* 12: 5- 7.
- Cazzola M, Donner C F, Hanania N A (2007). One hundred years of chronic pulmonary disease (COPD). *Respiratory medicine* 101: 1049-1065.
- Cooper JD, Patterson GA, Grossman R, Maurer J (1989) Double-lung transplant for advanced chronic obstructive lung disease. *Am Rev Respir Dis* 139: 303-307.
- Cooper JD, Trulock EP, Triantafyllou AN, Patterson GA, Pohl MS, et al. (1995) Bilateral pneumectomy (volume reduction) for chronic obstructive pulmonary disease. *J Thorac Cardiovasc Surg* 109: 106-116.
- Cotran R S, Kumar V, Robbins S L, Robbins J (1991) Patologia estrutural e funcional. 4.ed. Rio de Janeiro: Guanabara Koogan, p. 620-633.
- Dallegri F, Ottonello L (1997) Tissue injury in neutrophilic inflammation. *Inflamm Res* 46: 382-391.
- Datasus: Banco de Dados do Sistema Único de Saúde. Disponível em: Acesso em: 2013.
- Deslauries J (1995) History of Surgery for Emphysema, Seminar in Thorac and Cardiovasc Surg 8: 1
- Di Stefano A, Maestrelli P, Roggeri A, Turato G, Calabro S, et al. (1994) Upregulation of adhesion molecules in the bronchial mucosa of subjects with chronic obstructive bronchitis. *Am J Respir Crit Care Med* 149: 803-810.
- Di Stefano A, Capelli A, Lusuardi M, Balbo P, Vecchio C, et al. (1998) Severity of airflow limitation is associated with severity of airway inflammation in smokers. *Am J Respir Crit Care Med* 158: 1277-1285.
- Einhoven W (1995) Über die Wirkung der Bronchial-muskeln, nacheinerneun Methodeuntersucht, and Über Asthma nervosum. *Arch GesPhysiol* 51:367.
- Eriksson S (1965) Studies in alpha 1-antitrypsin deficiency. *Acta Med Scand Suppl* 432: 1-85.
- Holden C, Vogel G (2002) Stem cells. Plasticity: time for a reappraisal? *Science* 296: 2126-2129.

23. Huang Q, Liu D, Majewski P, Schulte LC, Korn JM, et al. (2001) The plasticity of dendritic cell responses to pathogens and their components. *Science* 294: 870-875.
24. Hyatt RE1, Farkas G, Schroeder M (2000) Pulmonary mechanics of papain emphysema in dogs *Chest* 117: 246S.
25. Ishizawa K, Kubo H, Yamada M, Kobayashi S, Numasaki M et al. (2004) Bone marrow-derived cells contribute to lung regeneration after elastase-induced pulmonary emphysema. *Federation of European Biochemical Societies* 556: 249-252.
26. Koca E, Champlin RE (2008) Peripheral blood progenitor cell or bone marrow transplantation: controversy remains. *Curr Opin Oncol* 20: 220-226.
27. Kotton DN, Ma BY, Cardoso WV, Sanderson EA, Summer RS, et al. (2001) Bone marrow-derived cells as progenitors of lung alveolar epithelium. *Development* 128: 5181-5188.
28. Kummel H (1923) Sr: Die operative heilungdes asthma bronchiale. *KlinWochenschr* 2:1825, 1923.
29. Laenec RTH (1819) Lauscultation mediate. Brasson JÁ and Chaude JS, Paris.
30. Lama VN , Smith L, Badri L, Flint A, Andrei AC, et al. (2007) Evidence for tissue-resident mesenchymal stem cells in human adult lung from studies of transplanted allografts. *J Clin Invest* 117: 989-996.
31. Hogg J C, Walker B A (1995) Polymorphonuclear leucocyte traffic in lung inflammation. *Thorax* 50: 819-820.
32. Laurell C B, Erickson S (1963) The electrophoretic alpha-1 deficiency. *Scand J Clin Lab. Invest* 15: 32-40.
33. Leuenberger P, Anderhub HP, Brändli O, Keller R, Knoblauch A, et al. (1997) Management 1997 of chronic obstructive pulmonary disease. Working Group of the Swiss Society of Pneumology. *Schweiz Med Wochenschr* 127: 766-782.
34. Nikula KJ , March TH, Seagrave J, Finch G, Barr E, et al. (2000) A mouse model of cigarette smoke-induced emphysema *Chest* 117: 246S-247S.
35. Orlic D, Kajstura J, Chimenti S, Limana F, Jakoniuk I, et al. (2001) Mobilized bone marrow cells repair the infarcted heart, improving function and survival. *Proc Natl Acad Sci U S A* 98: 10344-10349.
36. Pneumoatual Qual a frequência do enfisema pulmonar. Disponível em: Acesso em: 10 julho, 2013.
37. Prockop DJ (1997) Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 276: 71-74.
38. Prosper F E, Herreros J (2004) Células madre adultas. *Ver. Argent. Cardiol* 72: 68-73
39. Lagasse E , Connors H, Al-Dhalimy M, Reitsma M, Dohse M, et al. (2000) Purified hematopoietic stem cells can differentiate into hepatocytes in vivo. *Nat Med* 6: 1229-1234.
40. Krause DS, Theise ND, Collector MI, Henegariu O, Hwang S, et al. (2001) Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. *Cell* 105: 369-377.
41. Finkelstein R, Fraser RS, Ghezzi H, Cosio MG (1995) Alveolar inflammation and its relation to emphysema in smokers. *Am J Respir Crit Care Med* 152: 1666-1672.
42. Freund W A (1906) Zuroperativen Behandlunggewisser Lungenkraanheiteninsbesondere des auf starrer ThoraxdilatationberuhendenalveolareEmphysems. *Z ExpPatholTherap* 3:479.
43. Hellermann G R, Nagy S B, Kong, X.; Lockey, R.F.Mohapatra, S.S. (2002). Mechanism of cigarette smoke condensate-induced acute inflammatory response in human bronchial epithelial cells. *Respir Res* 3: 22.
44. Lopez AD, Murray CC (1998) The global burden of disease. 1990-2020. *Nat Med* 4: 1241-1243.
45. Mahadeva R , Shapiro SD (2002) Chronic obstructive pulmonary disease * 3: Experimental animal models of pulmonary emphysema. *Thorax* 57: 908-914.
46. Martin GR (1981) Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. *Proc Natl Acad Sci U S A* 78: 7634-7638.
47. Pushpakon R, Hogg J C, Woolcock A J, Angus A E, Macklem P T et al. (1970) Experimental papain-induced emphysema in dogs. *Am Rev Respir Dis* 102: 778-779.
48. Reich L (1924) Der einfluss des pneumoperitoneums auf das lungenemphysem. *Wien Arch Finn Med* 8:245-260.
49. Retamales I , Elliott WM, Meshi B, Coxson HO, Pare PD, et al. (2001) Amplification of inflammation in emphysema and its association with latent adenoviral infection. *Am J Respir Crit Care Med* 164: 469-473.
50. Rufino R, Lapa E Silva J R (2006) Bases celulares e bioquímicas da doença pulmonar obstrutiva crônica. *J Bras Penumol* 32: 241-248.
51. Russell RE , Thorley A, Culpitt SV, Dodd S, Donnelly LE, et al. (2002) Alveolar macrophage-mediated elastolysis: roles of matrix metalloproteinases, cysteine, and serine proteases. *Am J Physiol Lung Cell Mol Physiol* 283: L867-873.
52. Fusco L B, Pêgo-Fernandes, P M, Xavier A M, Pazetti R, Rivero D H R F (2002) Modelo experimental de enfisema pulmonar em ratos induzido por papaina. *J Penumol* 28: 1-7.
53. Gordon B (1934) The mechanism and use of abdominal supports and treatment of pulmonary diseases. *Am J Med Sci* 187:692-700.
54. Gross P, Pfitzer E A, Tolker E, Babyak M A, Kaschak M (1965) Experimental Emphysema: Its Production With Papain In Normal And Silicotic Rats. *Arch Environ Health* 11: 50-58.
55. Saetta M, Di Stefano A, Turato G, Facchini FM, Corbino L, et al. (1998) CD8+ T-lymphocytes in peripheral airways of smokers with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 157: 822-826.
56. Saetta M, Turato G, Maestrelli P, Mapp CE, Fabbri LM (2001) Cellular and structural bases of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 163: 1304-1309.
57. Schrepfer S, Deuse T, Reichenspurner H, Fischbein MP, Robbins RC, et al. (2007) Stem cell transplantation: the lung barrier. *Transplant Proc* 39: 573-576.
58. Senior R M, Anthonisen N R (1998) Chronic obstructive pulmonary disease (COPD). *Am J Resp Crit Care Med* 152: 5139-5147.
59. Shapiro SD (1994) Elastolytic metalloproteinases produced by human mononuclear phagocytes. Potential roles in destructive lung disease. *Am J Respir Crit Care Med* 150: S160-164.
60. Shapiro SD (2000) Evolving concepts in the pathogenesis of chronic obstructive pulmonary disease. In: C. Rochester ed. *Clinics in Chest Medicine* 633-644. Philadelphia, W.B. Saunders.

61. Suratt BT, Cool CD, Serls AE, Chen L, Varella-Garcia M, et al. (2003) Human pulmonary chimerism after hematopoietic stem cell transplantation. *Am J Respir Crit Care Med* 168: 318-322.
62. Takubo Y, Guerassimov A, Ghezzi H, Triantafyllou A, Bates JH, et al. (2002) Alpha1-antitrypsin determines the pattern of emphysema and function in tobacco smoke-exposed mice: parallels with human disease. *Am J Respir Crit Care Med* 166: 1596-1603.
63. Till JE, McCulloch EA (1961) A direct measurement of the radiation sensitivity of normal mouse bone marrow cells. *Radiat Res* 14: 213-222.
64. Turhan AG (2003) [Plasticity of adult stem cells]. *Transfus Clin Biol* 10: 103-108.
65. Van Bekkum DW (1966) Present status of bone marrow transplantation following whole body irradiation. *Oncology* 20: Suppl:60-72.
66. Verfaillie CM (2004) "Adult" Stem Cells: tissue Specific or Not ? In: *Handbook of Stem Cells. Vol 2. Adult and Fetal Stem Cells.* pp 1-20. Edit R. Lanza, H. Blau, D. Melton, M. Moore, D.D. Thomas, C. Verfaillie, I. Weissman and M. West . Elsevier Academic Press, UK.
67. Knudson R J, Gaensler E A (1965) Surgery For Emphysema. *Ann Thorac Surg* 122: 332-362.
68. Wagers AJ, Weissman IL (2004) Plasticity of adult stem cells. *Cell* 116: 639-648.
69. Wagner J G, Roth RA (2000) Neutrophil migration mechanisms, with an emphasis on the pulmonary vasculature. *Pharmacol Rev* 52: 349-374.
70. World Health Organization Strategy for prevention and control of chronic respiratory diseases WHO/MNC/CRA/02.1, 2002.
71. Yamada M, Kubo H, Kobayashi S, Ishizawa K, Numasaki M, et al. (2004) Bone marrow-derived progenitor cells are important for lung repair after lipopolysaccharide-induced lung injury. *J Immunol* 172: 1266-1272.
72. Zago M A, Covas D T (2006) Células-tronco: Origens e Propriedades. In: *Células-tronco: A nova Fronteira da medicina.* Ed. M. Zago e D. T. Covas, pp. 3-20. Editora Atheneu São Paulo.
73. Xu J, Qu J, Cao L, Sai Y, Chen C, et al. (2008) Mesenchymal stem cell-based angiopoietin-1 gene therapy for acute lung injury induced by lipopolysaccharide in mice. *J Pathol* 214: 472-481.
74. Carrell RW (2004) What we owe to alpha(1)-antitrypsin and to Carl-Bertil Laurell. *COPD* 1: 71-84.
75. Vidal R, Blanco I, Casas F, Jardí R, Miravittles M; Committee on the National Registry of Individuals with Alpha-1 Antitrypsin Deficiency (2006) [Guidelines for the diagnosis and management of alpha-1 antitrypsin deficiency]. *Arch Bronconeumol* 42: 645-659.
76. Costa X, Jardim R, Rodriguez F, Miravittles M, Cotrina M, et al. (2000) Simple method for alpha1-antitrypsin deficiency screening by use of dried blood spot specimens. *Eur Respir J* 15: 1111-1115.
77. Stoller JK, Aboussouan LS (2005) Alpha1-antitrypsin deficiency. *Lancet* 365: 2225-2236.
78. Seixas S, Garcia O, Trovoada MJ, Santos MT, Amorim A, et al. (2001) Patterns of haplotype diversity within the serpin gene cluster at 14q32.1: insights into the natural history of the alpha1-antitrypsin polymorphism. *Hum Genet* 108: 20-30.
79. de Serres FJ (2002) Worldwide racial and ethnic distribution of alpha1-antitrypsin deficiency: summary of an analysis of published genetic epidemiologic surveys. *Chest* 122: 1818-1829.
80. Menezes AM, Perez-Padilla R, Jardim JR, Muiño A, Lopez MV, et al. (2005) Chronic obstructive pulmonary disease in five Latin American cities (the PLATINO study): a prevalence study. *Lancet* 366: 1875-1881.
81. Hele D (2008) Meeting Report. First Siena International Conference on animal models of chronic obstructive pulmonary disease, Certosa di Pontignano, Universidade of Siena, Italy, September 30-October 2, 2001; *Respir Res* 3:12
82. de Faria EJ, de Faria IC, Alvarez AE, Ribeiro JD, Ribeiro AF, et al. (2005) [Association between alpha 1 antitrypsin deficiency and cystic fibrosis severity]. *J Pediatr (Rio J)* 81: 485-490.
83. Cruz PE, Mueller C, Cossette TL, Golant A, Tang Q, et al. (2007) In vivo post-transcriptional gene silencing of alpha-1 antitrypsin by adeno-associated virus vectors expressing siRNA. *Lab Invest* 87: 893-902.
84. Stockley RA, Luisetti M, Miravittles M, Piitulainen E, Fernandez P; Alpha One International Registry (AIR) group (2007) Ongoing research in Europe: Alpha One International Registry (AIR) objectives and development. *Eur Respir J* 29: 582-586.
85. Martorana PA, van Even P, Gardi C, Lungarella G (1989) A 16-month study of the development of genetic emphysema in tight-skin mice. *Am Rev Respir Dis* 139: 226-232.
86. Melton D A, Cowan C (2004) "Stemness": Definitions, criteria, and standards. In: Lanza, R. (editor-in-chief). *Handbook of Stem Cells. v. 1.* New York: Elsevier/Academic Press, p.xxv-xxxii.
87. Mendes R (1995) *Patologia do trabalho* 1 ed. Rio de Janeiro: Atheneu, p. 91-115.
88. Minguell JJ, Erices A, Conget P (2001) Mesenchymal stem cells. *Exp Biol Med (Maywood)* 226: 507-520.
89. Murray C J L, Lopez A D (1997) Mortality by cause for eight regions of the world: Global Burden of Disease Study. *Lancet* 349: 1269-1276.
90. Nakayama K (1961) Surgical removal of the carotid body for bronchial asthma. *Dis Chest* 40: 595-604.
91. National Institute Of Health (NHI) (2001). *Stem Cells: Scientific Progress and Future Research Directions.* June.
92. Neuringer IP, Randell SH (2004) Stem cells and repair of lung injuries. *Respir Res* 5: 6.
93. Russell RE, Culpitt SV, DeMatos C, Donnelly L, Smith M, et al. (2002) Release and activity of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 by alveolar macrophages from patients with chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol* 26: 602-609.
94. Lieberman J, Winter B, Sastre A (1986) Alpha 1-antitrypsin Pi-types in 965 COPD patients. *Chest* 89: 370-373.
95. Popescu LM, Fausone-Pellegrini MS (2010) TELOCYTES - a case of serendipity: the winding way from Interstitial Cells of Cajal (ICC), via Interstitial Cajal-Like Cells (ICLC) to TELOCYTES. *J Cell Mol Med* 14: 729-740.
96. Popescu LM, Manole CG, Gherghiceanu M, Ardelean A, Nicolescu MI, et al. (2010) Telocytes in human epicardium. *J Cell Mol Med* 14: 2085-2093.
97. Popescu L M (2011) Telocytes and stem cells: a tandem in cardiac regenerative medicine. *Curr Med Chem* 18: 94.

98. Popescu L M (2011) Telocytes- a novel type of interstitial cells. In: Braissant O, H Wakamatsu H, Kang I, et al, editors. Recent researches in modern medicine – HISTEM'11. Cambridge: WSEAS Press. p. 424–32.
99. Popescu L M, Gherghiceanu M, Kostin S, et al. (2011) Telocytes and heart renewing. *Adaptation biology and medicine*: 6: 17–39.
100. Popescu LM, Manole E, Serboiu CS, Manole CG, Suciuc LC, et al. (2011) Identification of telocytes in skeletal muscle interstitium: implication for muscle regeneration. *J Cell Mol Med* 15: 1379-1392.
101. Mandache E, Gherghiceanu M, Macarie C, Kostin S, Popescu LM (2010) Telocytes in human isolated atrial amyloidosis: ultrastructural remodelling. *J Cell Mol Med* 14: 2739-2747.
102. Gherghiceanu M, Popescu LM (2010) Cardiomyocyte precursors and telocytes in epicardial stem cell niche: electron microscope images. *J Cell Mol Med* 14: 871-877.
103. Faussone-Pellegrini MS, Popescu LM. (2011) Telocytes. *Biomol Concepts*;
104. Cantarero I, Luesma MJ, Junquera C (2011) The primary cilium of telocytes in the vasculature: electron microscope imaging. *J Cell Mol Med* 15: 2594-2600.
105. Cantarero Carmona I, Luesma Bartolomé MJ, Junquera Escribano C (2011) Identification of telocytes in the lamina propria of rat duodenum: transmission electron microscopy. *J Cell Mol Med* 15: 26-30.
106. Zheng Y, Cretoiu D, Yan G, Cretoiu SM, Popescu LM, et al. (2014) Protein profiling of human lung telocytes and microvascular endothelial cells using iTRAQ quantitative proteomics. *J Cell Mol Med* 18: 1035-1059.
107. Galiger C, Kostin S, Golec A, Ahlbrecht K, Becker S, et al. (2014) Phenotypical and ultrastructural features of Oct4-positive cells in the adult mouse lung. *J Cell Mol Med* 18: 1321-1333.
108. Zheng Y, Cretoiu D, Yan G, Cretoiu SM, Popescu LM, et al. (2014) Comparative proteomic analysis of human lung telocytes with fibroblasts. *J Cell Mol Med* 18: 568-589.
109. Zheng Y Bai C, Wang X (2012) Potential significance of telocytes in the pathogenesis of lung diseases. *Expert Rev Respir Med* 6: 45-49.
110. Popescu LM, Gherghiceanu M, Suciuc LC, Manole CG, Hinescu ME (2011) Telocytes and putative stem cells in the lungs: electron microscopy, electron tomography and laser scanning microscopy. *Cell Tissue Res* 345: 391-403.
111. Zheng Y, Li H, Manole CG, Sun A, Ge J, et al. (2011) Telocytes in trachea and lungs. *J Cell Mol Med* 15: 2262-2268.
112. Popescu LM, Gherghiceanu M, Suciuc LC, Manole CG, Hinescu ME (2011) Telocytes and putative stem cells in the lungs: electron microscopy, electron tomography and laser scanning microscopy. *Cell Tissue Res* 345: 391-403.
113. Hinescu ME, Gherghiceanu M, Suciuc L, Popescu LM (2011) Telocytes in pleura: two- and three-dimensional imaging by transmission electron microscopy. *Cell Tissue Res* 343: 389-397.
114. Suciuc LC, Popescu BO, Kostin S, Popescu LM (2012) Platelet-derived growth factor receptor- β^2 -positive telocytes in skeletal muscle interstitium. *J Cell Mol Med* 16: 701-707.
115. Díaz-Flores L, Gutiérrez R, García MP, Sáez FJ, Díaz-Flores L Jr, et al. (2014) CD34+ stromal cells/fibroblasts/fibrocytes/telocytes as a tissue reserve and a principal source of mesenchymal cells. Location, morphology, function and role in pathology. *Histol Histopathol* 29: 831-870.
116. Bani D, Formigli L, Gherghiceanu M, Faussone-Pellegrini MS (2010) Telocytes as supporting cells for myocardial tissue organization in developing and adult heart. *J Cell Mol Med* 14: 2531-2538.
117. Faussone-Pellegrini MS, Bani D (2010) Relationships between telocytes and cardiomyocytes during pre- and post-natal life. *J Cell Mol Med* 14: 1061-1063.
118. Luesma MJ, Gherghiceanu M, Popescu LM (2013) Telocytes and stem cells in limbus and uvea of mouse eye. *J Cell Mol Med* 17: 1016-1024.
119. Ceafalan L, Gherghiceanu M, Popescu LM, Simionescu O (2012) Telocytes in human skin--are they involved in skin regeneration? *J Cell Mol Med* 16: 1405-1420.
120. Manole C G, Cisma A, Yiu V, Gherghiceanu M, Popescu LM (2011) Experimental acute myocardial infarction: telocytes involvement in neo-angiogenesis. *J Cell Mol Med* 15: 2284-2296.